

James Bartley and Conroy Wong

Keywords

Airway inflammation • Nitric oxide • Carbon dioxide • Unified airway • Asthma • Chronic obstructive airway disease • Bronchiectasis • Allergic rhinitis • Chronic rhinosinusitis

Abbreviations

NO	Nitric oxide
CO ₂	Carbon dioxide
PaO ₂	Arterial oxygen levels
FEO ₂	Fraction of expired oxygen
FECO ₂	Fraction of expired carbon dioxide

Core Messages

- Physiological, epidemiological, and clinical evidence support an integrated upper and lower respiratory airway or “unified airway” model.

- Nasal breathing improves arterial oxygen concentrations and carbon dioxide excretion from the lungs.
- Nasal mucosal inflammation results in lower airway inflammation and vice versa.
- Many patients with asthma, chronic obstructive airway disease, and bronchiectasis have significant upper respiratory disease.
- Medical management of allergic rhinitis improves asthma.
- Nitric oxide from the nose and sinuses may have a role in the sterilization of incoming air and in improving ventilation-perfusion in the lungs.
- Nasal resistance is inversely related to end-tidal carbon dioxide levels.

J. Bartley, MB, ChB, FRACS, FFPMANZCA (✉)
Department of Otolaryngology –
Head and Neck Surgery,
Counties Manukau District Health Board,
19 Lambie Drive, Manukau, Auckland, New Zealand
e-mail: jbartley@ihug.co.nz

C. Wong, MBChB, Dip Obs, FRACP, CCST
Department of Respiratory Medicine,
Middlemore Hospital, 19 Lambie Drive,
Otahuhu, Auckland 1640, New Zealand
e-mail: cawong@middlemore.co.nz

37.1 Introduction

Physiological, epidemiological, and clinical evidence support an integrated upper and lower respiratory airway or “unified airway” model (Krouse et al. 2007; Guilemany et al. 2009; Hurst 2009; Marple 2010). Important, well-known nasal

functions include the filtering, warming, and humidification of inspired air before inhalation into the lungs. The nose and lungs potentially interact with each other in a number of ways (Fig. 37.1). Nasal mucosal inflammation results in lower airway inflammation and vice versa (Braunstahl et al. 2001a, b). Part of this mechanism is thought to be a generalized inflammatory response that amplifies the response to inflammatory stimuli in other parts of the respiratory tract (Braunstahl et al. 2001a, b). Inflammatory mediators and/or infectious pathogens may also be carried along the respiratory mucosa or along air currents (Hare et al. 2010). Neuronal responses may play a role, although the existence of nasobronchial reflexes remains controversial (Sarin et al. 2006). Nitric oxide (NO) from the nose and sinuses may have a role in the sterilization of incoming air

(Lundberg et al. 1995) and in improving ventilation-perfusion in the lungs (Selimoglu 2005). An inverse relationship between nasal resistance and end-tidal carbon dioxide (CO_2) levels has been described (Mertz et al. 1984; Shi et al. 1988). CO_2 and NO may act as aerocrine messengers (Bartley 2005; Selimoglu 2005). Olfaction is also linked to the limbic system (Soudry et al. 2011), which can independently control our breathing pattern and rate (Plum 1992).

Nasal breathing improves arterial oxygen concentrations and carbon dioxide excretion from the lungs.

37.2 Physiological Interactions

The nose has an important role in filtering, warming, and humidifying inspired air before inhalation into the lungs. The nose also provides a resistance to both inspiration and expiration that is twice that of the open mouth. This increased resistance appears to have a number of physiological benefits. In a study of arterial oxygen levels (PaO_2) before and after jaw wiring, which forced patients to breathe continuously through their noses, PaO_2 increased by nearly 10 % (Swift et al. 1988). Nasal packing after nasal surgery forcing patients to breathe through their mouths is associated with a reduction in arterial O_2 saturation (Ogretmenoglu et al. 2002). At rest, end-tidal CO_2 levels in expired air, which are considered to be a reliable indirect measure of CO_2 levels in the arterial blood, increase with nasal breathing indicating that nasal breathing improves the efficiency of CO_2 excretion from the lungs (Tanaka et al. 1988). During exercise, nasal breathing reduces the fraction of expired oxygen ($F_{\text{E}\text{O}_2}$), indicating that on expiration the percentage of O_2 extracted from the air by the lungs is increased, and increases the fraction of expired carbon dioxide ($F_{\text{E}\text{CO}_2}$), indicating an increase in the percentage of expired air that is CO_2 (Morton et al. 1995). This equates to more efficient O_2 extraction and CO_2 excretion during exercise.

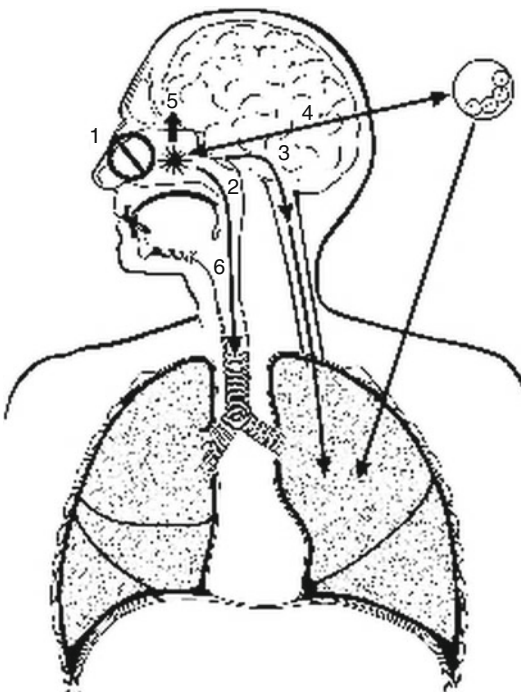


Fig. 37.1 Potential mechanisms for interaction between the upper and lower airways: (1) loss of nasal conditioning function, (2) direct passage of inflammatory mediators and/or microorganisms between the upper and lower airways, (3) nasobronchial reflexes, (4) stimulation at one point of the respiratory mucosal surface results in a pan-airway inflammatory response, (5) olfactory regulation of respiration, and (6) aerocrine messengers (CO_2 and NO)

Explanations for these observations still remain largely hypothetical. Nasal breathing increases total lung volume (Swift et al. 1988). The corresponding increase in functional residual capacity (volume of air present in the lungs present after passive expiration) is thought to improve gas exchange leading to improved PaO₂. NO derived from the nose and sinuses might also improve ventilation-perfusion relationships in the lungs (Della Rocca and Coccia 2005). The nose also provides an inspiratory resistance forcing the diaphragm to contract against a resistance. On a long-term basis, this might also have an important role in maintaining diaphragmatic muscle strength, although the role of inspiratory muscle training in a range of lung diseases continues to be debated (Padula and Yeaw 2007; Gosselink et al. 2011). Regardless, nasal breathing appears important in aiding O₂ absorption and in facilitating CO₂ excretion in the lungs.

Nasal breathing significantly improves oxygen extraction and carbon dioxide excretion during exercise.

The breathing cycle is divided into inspiratory and expiratory phases. When the respiratory rate increases, the expiratory phase shortens. Nasal breathing slows the respiratory rate increasing the length of the expiratory phase (Ayoub et al. 1997). Increasing the expiratory phase of the respiratory cycle is known to increase the body's relaxation response (Cappo and Holmes 1984). A number of ancient disciplines, such as yoga and tai chi, emphasize the importance of nasal breathing in relaxation and meditation.

37.3 Respiratory Inflammation

Upper and lower airway inflammatory processes often coexist and share common pathogenic mechanisms (Selimoglu 2005; Hare et al. 2010; Marple 2010). Based on the predominant cell type, chronic rhinosinusitis (CRS) has been

classified as being either eosinophilic or neutrophilic (Meltzer and Hamilos 2011). The eosinophilic group includes CRS with polyps, a subset of CRS without polyps, aspirin hypersensitivity, asthma and nasal polyps (Samter's triad), and allergic fungal rhinosinusitis. Eosinophilic CRS and allergic rhinitis are associated with asthma (Marple 2010). Lower airway inflammation has also been classified according to the cell profile of induced sputum as being either eosinophilic or non-eosinophilic (Hargreave 2007). In asthma patients, eosinophils are the dominant inflammatory cells in middle meatal lavage (Ragab et al. 2005). In small airway disease patients, neutrophils are the dominant inflammatory cells in middle meatal lavage (Ragab et al. 2005).

Asthma and allergic rhinitis are strongly inter-related (Corren 1997; Krouse et al. 2007; Marple 2010). Corren reported nasal symptoms in 78 % of asthmatic patients and that 38 % of allergic and nonallergic rhinitis patients have asthma (Corren 1997). The severity of asthma symptoms correlates closely with rhinitis symptoms (Krouse et al. 2007). The presence of allergic rhinitis also increases the risk of subsequent asthma development nearly fourfold (Shaaban et al. 2008).

Asthma has also been associated with CRS. The prevalence of asthma is 20 % in CRS patients, which is higher than that seen in the general population (Jani and Hamilos 2005). Asthma severity also correlates with CRS disease severity, as determined by computed tomography (Bresciani et al. 2001). In patients having functional endoscopic sinus surgery, the asthma prevalence is 42 %, rising to 50 % in those with nasal polyps (Senior et al. 1999).

Neutrophilic inflammation is a feature of both chronic obstructive pulmonary disease (COPD) and bronchiectasis (Hargreave 2007). While tissue eosinophilia is an established feature of asthma, a neutrophilic picture can also be seen (Hargreave 2007). In COPD patients, inflammatory cells are found both in the sputum and in lung biopsy specimens (Hurst 2009). COPD patients commonly report nasal symptoms, the most common of which is rhinorrhea (Hurst et al. 2006). Nasal symptoms are also more common in COPD patients with chronic sputum production

(Hurst et al. 2006). Nasal symptoms double the risk over 8 years of patients developing COPD (Nihlén et al. 2008). Increased levels of the neutrophil chemoattractant protein IL-8 are found in the upper airways of COPD patients, when compared to control subjects (Hurst et al. 2006). Upper airway IL-8 concentrations correlate with those in the lower airway, and the concentration at both sites is related to indexes of bacterial colonization (Hurst et al. 2006). Many bronchiectatic patients also have nasal and sinus disease. Most bronchiectasis patients (77 %) meet the diagnostic criteria for CRS with 25 % of bronchiectasis patients having nasal polyps. Bronchiectasis severity also correlates with CRS severity (Guilemany et al. 2009).

Patients with asthma, chronic obstructive pulmonary disease, and bronchiectasis frequently have upper respiratory disease.

37.3.1 Inflammatory Interactions

Under the influence of serum IL-5 and eotaxin, eosinophils are released from the bone marrow into the systemic circulation. Depending on the local expression of a variety of adhesion molecules, cytokines, and chemokines, eosinophils then migrate to inflamed areas. Leukocytes migrate along a chemotactic gradient through the endothelium. Local cells upregulate endothelial adhesion molecules through the release of IL-1 β , IL-4, and TNF- α . Leukocyte endothelial adherence is increased. Intercellular adhesion molecule 1, vascular cell adhesion molecule 1, and E-selectin are known endothelial adhesion molecules (Kita 2011). Increased local expression of these adhesion molecules occurs after experimental nasal and bronchial allergen challenge. Experimentally this results in increased eosinophilic allergic inflammation in the nasal and bronchial mucosa (Braunstahl et al. 2001a, b).

Even a single nasal allergen challenge administered to non-asthmatic subjects with seasonal allergy increases blood eosinophil levels and IL-5 in both the upper and lower airways (Corren 1997). Prior nasal stimulation using a nasal provocation

antigen challenge increases bronchial hyperresponsiveness to methacholine challenge (Bonay et al. 2006). Similarly segmental bronchial provocation in patients with allergic rhinitis induces blood eosinophilia and mucosal inflammation in both the upper and lower airways. This inflammation is characterized by increased numbers of eosinophils, IL-5+ cells, and eotaxin-positive cells. Local allergen exposure in both the upper and lower airways results in generalized airway inflammation – this would appear to occur through vascular mechanisms.

A high level of similarity in bacterial cultures from the nasopharynx and from bronchoalveolar lavage is seen in children with bronchiectasis and protracted bacterial bronchitis as well as in patients with cystic fibrosis.

37.3.2 Microbial Aspiration

The silent aspiration of nasopharyngeal secretions has been hypothesized as important in relationships between upper and lower respiratory airway disease (Bardin et al. 1990; Kogahara et al. 2009). The possibility that *Staphylococcus aureus*-derived enterotoxins could also be inhaled into the lower airway has been raised (Hamilos 2000). COPD patients with lower airway bacterial colonization have a higher total nasal bacterial load (Hurst et al. 2005). Children with bronchiectasis have a high nasopharyngeal carriage of *Streptococcus pneumoniae*, nontypable *Haemophilus influenzae* (NTHi), and *Moraxella catarrhalis* and lower airway infection by NTHi. A high level of similarity in cultures from the nasopharynx and from bronchoalveolar lavage is seen (Hare et al. 2010). Similar agreement is also seen in children with protracted bacterial bronchitis (Hare et al. 2010) and in patients with cystic fibrosis (Godoy et al. 2011). In the critical care situation, the treatment of upper respiratory disease reduces the risk of ventilator-associated pneumonia (de Smet et al. 2009).

People with a high nasal bacterial load (and associated greater nasal inflammation) appear

more likely to pass bacteria into the lower respiratory tract. The evidence that gross aspiration of nasopharyngeal secretions into the lower respiratory airway occurs is controversial. Radionucleotide scanning in humans shows that it does not occur (Bonay et al. 2006); however, animal experiments suggest that it may occur during sleep (Kogahara et al. 2009). The similarity in microbiological cultures between the upper and lower respiratory tract suggests that some transmission occurs. Microaspiration of bacteria and inflammatory mediators could possibly occur, but this has yet to be demonstrated scientifically.

37.3.3 Influence of Upper Respiratory Interventions on the Lower Respiratory Tract

In the majority of trials, the treatment of allergic rhinitis with nasal steroids reduces asthma severity (Stelmach et al. 2005; Krouse et al. 2007). At the end of 3 years, the children treated with immunotherapy for grass and/or birch allergic rhinoconjunctivitis were less likely than the control group to develop asthma (Möller et al. 2002). Similarly, a number of studies have shown that the surgical treatment of CRS helps asthma patients through both an improvement in asthma symptoms and the decreased use of asthma medication (Krouse et al. 2007). Unfortunately these studies have been uncontrolled. There are also no randomized controlled studies that have investigated the effect of CRS medical therapy on asthma (Krouse et al. 2007).

Medical and surgical treatment of upper respiratory disease helps asthmatic patients.

37.4 Nasobronchial Reflexes

Nasobronchial reflexes have also been implicated in the interactions between the upper and lower respiratory airways. Mechanical or chemical stimulation of nasal, tracheal, and laryngeal receptors

could produce sneezing, coughing, or bronchoconstriction, thus preventing deeper penetration of allergens or irritants into the airway (Sarin et al. 2006). Unilateral models of nasal provocation show that secretory responses can be measured in both nostrils (Sarin et al. 2006). The mechanism appears to be neural (Sarin et al. 2006).

When asthmatic patients exercise with their noses occluded, a 20 % decline in forced expiratory flow occurs, compared with less than a 5 % reduction among patients allowed to exercise while nose breathing (Shturman-Ellstein et al. 1978). Recent research has shown that bursts of cold air on the nasal mucosa increase nasal resistance. This effect was blocked by anesthetizing the nose or by inhaling atropine, an anticholinergic drug, before the provocation (Fontanari et al. 1996, 1997). Other researchers have not been able to confirm these results (Johansson et al. 2000). The existence of nasobronchial reflexes secondary to inflammatory exposure in the upper respiratory airway remains controversial (Sarin et al. 2006).

37.5 Olfaction and the Limbic System

Evolutionary theory teaches that in primitive life forms, the olfactory brain was probably a layer of cells above the brain stem that registered a smell and then simply categorized it. New layers of the olfactory brain then developed into what was initially called the rhinencephalon (nose brain) or limbic system (Le Doux 2003). Our limbic system coordinates the stress “fight or flight” response. As part of this response, not only does the heart rate increase, but the respiratory rate also goes up; the limbic system is able to override normal $p\text{CO}_2$ homeostasis (Plum 1992). The physiological evidence suggests that fragrances such as rosemary and lavender may have direct effects on human memory and mood (Herz 2009). Fragrances such as lavender by influencing the limbic system and the stress response could potentially affect breathing patterns and rate. The pheromone androstadienone when applied to women’s nasovomerine organs slowed both their heart and breathing rates (Grosser et al. 2000).

37.6 Aerocrine Communication

37.6.1 Nitric Oxide

NO is a gas that is produced by the nose and paranasal sinuses. NO has bacteriostatic actions (Lundberg et al. 1995). NO may have a role in the sterilization of incoming air (Lundberg et al. 1995) and in improving ventilation-perfusion in the lungs (Selimoglu 2005).

37.6.2 Carbon Dioxide

An inverse relationship between nasal resistance and end-tidal $p\text{CO}_2$ levels has been described (Mertz et al. 1984; Shi et al. 1988). A reduction of end-tidal expired $p\text{CO}_2$ from 40 to 35 mmHg (5.3 to 4.7 kPa) corresponds to an increase in nasal resistance of 10 % (Mertz et al. 1984). Breathing is commonly taught as being controlled by independent voluntary and metabolic pathways. However, the limbic system is able to override metabolic respiratory control systems (Plum 1992). People practicing yoga would appear to set their $p\text{CO}_2$ receptors to a higher response level. End-tidal $p\text{CO}_2$ concentrations are nearly 4 mmHg higher in yogic breathers (Stanescu et al. 1981). In contrast, people who

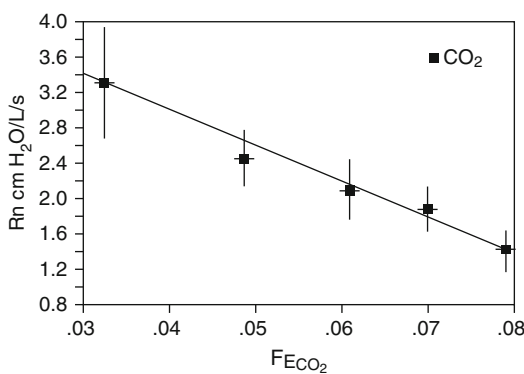


Fig. 37.2 Inverse linear relationship of nasal resistance to end-tidal expiratory CO_2 concentrations. An end-tidal $p\text{CO}_2$ reading of 0.05 corresponds to 38 mmHg. A reduction of end-tidal expired $p\text{CO}_2$ from 40 (0.053 kPa) to 35 mmHg (0.046 kPa) corresponds to an increase in nasal resistance of 10 % (Reproduced with permission from Mertz et al. 1984)

are prone to anxiety attacks would appear to have lower arterial $p\text{CO}_2$ levels (5 mmHg on average) as compared with controls (Papp et al. 1989). The mechanisms of the relationship between expired $p\text{CO}_2$ levels and nasal resistance are unknown. This could be either a systemic vascular effect or an aerocrine effect. Regardless, people who practice relaxed diaphragmatic breathing may be less likely to present complaining of nasal congestion, whereas anxious people are (Bartley 2006) (Fig. 37.2).

Conclusions

From a physiological perspective the nose has an important role in the preparation of inspired air before inhalation into the lungs. Other physiological effects such as improvements in O_2 transfer and CO_2 excretion would also appear to occur. Inflammation, both allergic and infective, affects both the upper and lower respiratory airways. These interactions would appear to occur through vascular mechanisms.

Both medical and surgical treatments of upper respiratory disease appear to help asthmatic patients. Both NO and CO_2 would appear to have roles in aerocrine communication. The influence of nasobronchial reflexes and microaspiration remains largely hypothetical. The current evidence indicates that optimal management of disease processes in both the upper and lower respiratory airways needs to consider a “unified airway” model.

References

- Ayoub J, Cohendy R, Dauzat M, Targhetta R, De la Coussaye J, Bourgeois J, et al. Non-invasive quantification of diaphragm kinetics using m-mode sonography. *Can J Anaesth.* 1997;44:739–44.
- Bardin P, Van Heerden B, Joubert J. Absence of pulmonary aspiration of sinus contents in patients with asthma and sinusitis. *J Allergy Clin Immunol.* 1990;86:82–3.
- Bartley J. Nasal congestion and hyperventilation syndrome. *Am J Rhinol.* 2005;19:607–11.
- Bartley J. Nasal congestion and hyperventilation syndrome. *Am J Rhinol.* 2006;19:607–11.
- Bonay M, Neukirch C, Grandsaigne M, Leçon-Malas V, Ravaud P, Dehoux M, et al. Changes in airway inflam-

- mation following nasal allergic challenge in patients with seasonal rhinitis. *Allergy*. 2006;61:111–8.
- Braunstahl G, Overbeek S, Fokkens W, Kleinjan A, McEuen A, Walls A, et al. Segmental bronchoprovocation in allergic rhinitis patients affects mast cell and basophil numbers in nasal and bronchial mucosa. *Am J Respir Crit Care Med*. 2001a;164:858–65.
- Braunstahl G, Overbeek S, KleinJan A, Prins J-B, Hoogsteden H, Fokkens W. Nasal allergen provocation induces adhesion molecule expression and tissue eosinophilia in upper and lower airways. *J Allergy Clin Immunol*. 2001b;107:469–76.
- Bresciani M, Paradis L, Des Roches A, Vernhet H, Vachier I, Godard P, et al. Rhinosinusitis in severe asthma. *J Allergy Clin Immunol*. 2001;107:73–80.
- Cappo B, Holmes D. The utility of prolonged respiratory exhalation for reducing physiological and psychological arousal in non-threatening and threatening situations. *J Psychosom Res*. 1984;28:265–73.
- Corren J. Allergic rhinitis and asthma: how important is the link? *J Allergy Clin Immunol*. 1997;99:S781–6.
- De Smet A, Kluytmans T, Cooper B, Mascini EM, Benus RF, van der Werf TS, et al. Decontamination of the digestive tract and oropharynx in ICU patients. *N Engl J Med*. 2009;360:20–31.
- Della Rocca G, Coccia C. Nitric oxide in thoracic surgery. *Minerva Anesthesiol*. 2005;71:313–8.
- Fontanari P, Burnet H, Zattara-Hartman M, Jammes Y. Changes in airway resistance induced by nasal inhalation of cold dry, or moist air in normal individuals. *J Appl Physiol*. 1996;81:1739–43.
- Fontanari P, Zattara-Hartmann M-C, Burnet H, Jammes Y. Nasal eupnoic inhalation of cold, dry air increases airway resistance in asthmatic patients. *Eur Respir J*. 1997;10:2250–4.
- Godoy J, Godoy A, Ribalta G, Largo I. Bacterial pattern in chronic sinusitis and cystic fibrosis. *Otolaryngol Head Neck Surg*. 2011;145:673–6.
- Gosselink R, De Vos J, van den Heuvel S, Segers J, Decramer M, Kwakkel G. Impact of inspiratory muscle training in patients with COPD: what is the evidence? *Eur Respir J*. 2011;37:416–25.
- Grosser B, Monti-Bloch L, Jennings-White C, Berliner D. Behavioral and electrophysiological effects of androstadienone, a human pheromone. *Psychoneuroendocrinology*. 2000;25:289–99.
- Guilemany JM, Angrill J, Alobid I, Centellas S, Pujols L, Bartra J, et al. United airways again: high prevalence of rhinosinusitis and nasal polyps in bronchiectasis. *Allergy*. 2009;64:790–7.
- Hamilos D. Chronic sinusitis. *J Allergy Clin Immunol*. 2000;106:213–27.
- Hare KM, Grimwood K, Leach AJ, Smith-Vaughan H, Torzillo PJ, Morris PS, et al. Respiratory bacterial pathogens in the nasopharynx and lower airways of Australian indigenous children with bronchiectasis. *J Pediatr*. 2010;157:1001–5.
- Hargreave F. Quantitative sputum cell counts as a marker of airway inflammation in clinical practice. *Curr Opin Allergy Clin Immunol*. 2007;7:102–6.
- Herz R. Aromatherapy facts and fictions: a scientific analysis of olfactory effects on mood, physiology and behavior. *Int J Neurosci*. 2009;119:263–90.
- Hurst JR. Upper airway. 3: Sinonasal involvement in chronic obstructive pulmonary disease. *Thorax*. 2009;65:85–90.
- Hurst JR, Wilkinson TM, Perera WR, Donaldson GC, Wedzicha JA. Relationships among bacteria, upper airway, lower airway and systemic inflammation in COPD. *Chest*. 2005;127:1219–26.
- Hurst J, Perera W, Wilkinson T, Donaldson G, Wedzicha J. Systemic and upper and lower airway inflammation at exacerbation of chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. 2006;173:71–8.
- Jani A, Hamilos D. Current thinking on the relationship between rhinosinusitis and asthma. *J Asthma*. 2005;42:1–7.
- Johansson A, Bende M, Millqvist E, Bake B. Nasobronchial relationship after cold air provocation. *Respir Med*. 2000;94:1119–22.
- Kita H. Eosinophils: multifaceted biological properties and roles in health and disease. *Immunol Rev*. 2011;242:161–77.
- Kogahara T, Kanai K, Asano K, Suzaki H. Evidence for passing down of postnasal drip into respiratory organs. *In Vivo*. 2009;23:297–301.
- Krouse J, Brown R, Fineman S, Han J, Heller A, Joe S, et al. Asthma and the unified airway. *Otolaryngol Head Neck Surg*. 2007;136:S75–106.
- Le Doux J. *The emotional brain*. New York: Phoenix; 2003.
- Lundberg J, Farkas-Szallasi T, Weitzberg E, Rinder J, Lidholm J, Anggård A, et al. High nitric oxide production in human paranasal sinuses. *Nat Med*. 1995;1:370–3.
- Marple BF. Allergic rhinitis and inflammatory airway disease: interactions within the unified airspace. *Am J Rhinol Allergy*. 2010;24:249–54.
- Meltzer E, Hamilos D. Rhinosinusitis diagnosis and management for the clinician: a synopsis of recent consensus guidelines. *Mayo Clin Proc*. 2011;86:427–43.
- Mertz J, McCaffrey T, Kern E. Role of the nasal airway in regulation of airway resistance during hypercapnia and exercise. *Otolaryngol Head Neck Surg*. 1984;92:302–7.
- Möller C, Dreborg S, Ferdousi HA, Halken S, Høst A, Jacobsen L, et al. Pollen immunotherapy reduces the development of asthma in children with seasonal rhinoconjunctivitis (the PAT study). *J Allergy Clin Immunol*. 2002;109:251–6.
- Morton A, King K, Papalia S, Goodman C, Turley K, Wilmore J. Comparison of maximal oxygen consumption with oral and nasal breathing. *Aust J Sci Med Sport*. 1995;27:51–5.
- Nihlén U, Montnémerly P, Andersson M, Persson CG, Nyberg P, Löfdahl CG, et al. Specific nasal symptoms

- and symptom producing factors may predict increased risk of developing COPD. *Clin Physiol Funct Imaging*. 2008;28:240–50.
- Ogretmenoglu O, Yilmaz T, Rahimi K, Aksöyek S. The effect on arterial blood gases and heart rate of bilateral nasal packing. *Eur Arch Otorhinolaryngol*. 2002; 259:63–6.
- Padula C, Yeaw E. Inspiratory muscle training: integrative review of use in conditions other than COPD. *Res Theory Nurs Pract*. 2007;21:98–118.
- Papp L, Martinez J, Klein D, Ross D, Liebowitz M, Fyer A, et al. Arterial blood gas changes in panic disorder and lactate-induced panic. *Psychiatry Res*. 1989;28(2):171–80.
- Plum F. Breathing is controlled independently by voluntary, emotional and metabolically related pathways. *Arch Neurol*. 1992;49:441.
- Ragab A, Clement P, Vincken W. Correlation between the cytology of the nasal middle meatus and BAL in chronic rhinosinusitis. *Rhinology*. 2005;43:11–7.
- Sarin S, Udem B, Sanico A, Togias A. The role of the nervous system in rhinitis. *J Allergy Clin Immunol*. 2006;118:999–1014.
- Selimoglu E. Nitric oxide in health and disease from the point of view of the otorhinolaryngologist. *Curr Pharm Des*. 2005;11:3051–60.
- Senior B, Kennedy DW, Tanabodee J. Long-term impact of functional endoscopic sinus surgery of asthma. *Otolaryngol Head Neck Surg*. 1999;121:66–8.
- Shaaban R, Zureik M, Soussan D, Neukirch C, Heinrich J, Sunyer J, et al. Rhinitis and the onset of asthma: a longitudinal population based study. *Lancet*. 2008;372:1049–57.
- Shi Y-X, Seto-Poon M, Wheatley J. Alae nasi activation decreases nasal resistance during hyperoxic hypercapnia. *J Appl Physiol*. 1988;85:294–300.
- Shturman-Ellstein R, Zeballos R, Buckley J, Souhrada J. The beneficial effect of nasal breathing on exercise-induced bronchoconstriction. *Am Rev Respir Dis*. 1978;118:65–73.
- Soudry Y, Lemogne C, Malinvaud D, Consoli S, Bonfils P. Olfactory system and emotion: common substrates. *Eur Ann Otorhinolaryngol Head Neck Dis*. 2011;128:18–23.
- Stanescu D, Nemery B, Verityer C, Marechal C. Pattern of breathing and ventilatory response to CO₂ in subjects practicing hatha yoga. *J Appl Physiol*. 1981;51:1625–9.
- Stelmach R, do Patrocínio T, Nunes M, Ribeiro M, Cukier A. Effect of treating allergic rhinitis with corticosteroids in patients with mild-to moderate persistent asthma. *Chest*. 2005;128:3140–7.
- Swift A, Campbell I, McKown T. Oronasal obstruction, lung volumes, and arterial oxygenation. *Lancet*. 1988; 1:73–5.
- Tanaka Y, Morikawa T, Honda Y. An assessment of nasal functions in control of breathing. *J Appl Physiol*. 1988; 65:1520–4.